



Aerobic

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A CULTURE SET CONSISTS OF TWO BACTEC CULTURE BOTTLES

- One aerobic (blue, collected 1st) and one anaerobic (purple, collected 2nd)

PREPARATION TIPS

- Visually inspect bottles for contamination, cracks or signs of deterioration
- Check the expiration date
- The recommended fill volume for each BACTEC culture bottle is 8-10mL
 - Using a syringe for collection helps ensure correct volume draw
 - Alternatively, using media meniscus as a guide, mark the optimal fill volume on each bottle (each line mark is approx. 5mL)



CAUTION: Underfilling or overfilling reduces sensitivity which may adversely affect recovery of microorganisms

COLLECTION & PROCESSING

1. Skin Preparation

- Cleanse venipuncture site for 30 seconds with a 70% alcohol prep pad and another 30 seconds using iodine (e.g., Betadine)
- Adequate disinfection is mandatory to minimize contamination from skin and environmental organisms

2. Preparation of BACTEC Culture Bottles

- Remove flip-off caps from culture bottles
- DISINFECT RUBBER BOTTLE TOPS WITH A 70% ALCOHOL PREP PAD AND ALLOW TO DRY IMMEDIATELY PRIOR TO COLLECTION
- Do not use any form of iodine to disinfect the tops of the bottles. It may affect the integrity of the rubber top of the culture bottle, causing false positive results

3. Collection

- Push and hold Vacutainer holder over top of aerobic bottle to puncture septum
- Collect recommended fill volume on the aerobic bottle. Monitor to ensure proper flow and fill level
- Remove Vacutainer holder from aerobic bottle. Immediately push and hold onto anaerobic bottle
- Collect blood or fluid to recommended fill volume on the anaerobic bottle. Remove Vacutainer holder from bottle and discard

4. Labeling and Shipping

- Bottles should be properly labeled with all required patient information
- Do not cover the BACTEC culture bottle barcode with patient's barcode label
- Label each culture set with a separate order number, and place in a transport bag
- Blood and fluid cultures should be held and shipped at room temperature, not refrigerated
- Specimens should be received no later than **48 hours** after collection to ensure optimal recovery of microorganisms